MOLECULAR IDENTIFICATION OF Salmonella Enterica FROM PATIENTS WITH DIARRHEA IN DUHOK GOVERNORATE KURDISTAN REGION / IRAQ.

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ABSTRACT

Background and objectives: Salmonella enterica are responsible for causing the highest number of bacterial foodborne infections in the world. Methods: The Present study was carried out for detection and molecular identification of Salmonella enterica isolated from stool specimens of patients suffering from diarrhoea using traditional method and identification by using of RT-Polymerase Chain Reaction (PCR) techniques. Results: Out of the 121 patients with diarrhoea attended Azadi and Heevi Teaching Hospitals, 72 cases (59.504%) gave positive results for culturing. While by PCR method, 68 cases (56.198%) gave positive results for Salmonella enterica. In this study, (49) cases (40.495%) of non-Salmonella spp. gave negative results by culture and (53) cases (43.8%) of non-Salmonella spp. gave negative results by PCR method. Conclusions: The current study showed that RT-PCR technique is more accurate and sensitive compared to traditional methods.

KEYWORD: Salmonella enterica, Food borne infections and RT-PCR.

INTRODUCTION

Salmonella are Gram-negative bacilli and non-spore forming, facultative anaerobic, non-lactose fermenting bacteria within the family of enterobacteriaceae. Infection with Salmonella is the most frequent food-borne gastrointestinal disease transmitted from animal products to human by food mainly through water, meat, eggs and poultry (Riyaz-Ul-Hassan, et al., 2004). Salmonella infection remains as a serious problem to public health which in turn is regarded as a major public health concern and responsibility for the significant cost in all around the world which leads to the thousands of deaths (Lynch, et al., 2006).

WHO estimated that the annual median number of non-typhoidal salmonellosis was 78.7 million foodborne illnesses with over 59 000 deaths, (Havelaar AH *et al.*, 2010). Non-typhoidal Salmonella species are a common cause of foodborne disease in the Middle East and North Africa (MENA). Moreover, *Salmonella* has been detected in an array of food products presented to

consumers in the region. The number and quality of the studies differ substantially by country. To the best of our knowledge, there has been no published study that systematically reviewed, synthetized and assessed the available data on non-typhoidal enteric *Salmonella* in human and food in the MENA region (Havelaar AH *et al.*, 2010).

There are many tools for diagnosis of salmonella infection: traditional bacteriological methods offered standardized procedures for microbial detection. However, they are time consuming that take approximately 4–7 days (Ferre et al., 2002) and not always compatible with short-time-to result demand. Therefore, Microbiology aims for supplementation of classical methods with molecular techniques based on detection of the microbial nucleic acids, which shorten the analysis time and lower the limit of detection. An important aspect in characterization of bacteria is the molecular detection in determining the clonal and strain distributions various environments. among Traditional microbial methods, albeit generally considered to be variable, labor-intensive, and time-consuming, are of practical value in epidemiological investigations. Molecular detection methods are mainly based on the analysis of the genetic material of microbial agents (Ranjbar R, et al., 2014, Lin T, et al., 2014). This study was undertaken to determine the presence, of salmonella bacteria from patients with diarrhea using traditional, biochemical and Molecular tools in Duhok governorate, Kurdistan Region of Iraq.

PATIENTS AND METHODS

Salmonella isolates were collected from Azadi Teaching Hospital at emergency department and Heevi Pediatric Hospital in Duhok City. These isolates were obtained from stools samples taken from 121 patients with salmonellosis or suspicious salmonellosis were diagnosed in outpatient clinics at Azadi Teaching Hospital and in Duhok Medical Research Center (DMRC) during April to October 2018.

Stools samples were pre-enriched in peptone water in sterile conditions. Depending on reproduction abilities of colonies and incubated at 37°c for 16-18 Hrs., enriched in Rappaport

Vassiliadis(VRS) at 42°c for overnight incubation and cultured on XLD agar according to International Organization for Standardization (ISO 6579-1:2017) then suspicious colonies taken for biochemical tests. DNA was extracted from 1 ml of the pre-enrichment broths using QIAamp DNA Extraction Mini Kit (Qiagen, Germany) following the manufacturers' instructions.

Polymerase Chain Reaction (PCR) was performed depending on amplification of the *invA* gene. The Final volume of amplification was 20µl, which described in Table (A-1). The PCR reaction consisted of 2X master mix and probed primer (10µl of Master Mix, 10µl of Extracted DNA).

The cycling parameters consisted of an initial PCR activation step, activation of HotStarTaq Plus DNA Polymerase (5 min, 95 °C), followed by 40×3step-cycles consisting of denaturation (15 sec. at 95°C), annealing for 15 sec. at 60 °C, and extension for 10 sec. at 72 °C, as in Table (A-2). Fluorescence detection was performed at the end of the annealing stage of each cycle. The amplification done by Rotor-gene Q instrument in Central Laboratory —Duhok.

Component Sample Positive PCR control Negative PCR control Reconstituted mericon 10 μΙ 10 µl 10 µl Sample DNA 10 μΙ Dissolved Positive Control DNA 10 µl QuantiTect Nucleic Acid Dilution Buffer or RNase-free 10 µl water Total volume 20 µl 20 µl 20 µl

Table A.1: Components of PCR reaction.

Table A-2: PCR program of Cycling parameters.

Step	Time	Temperature	Comments	
Initial PCR activation step	5 min	95℃	Activation of HotStarTaq Plu DNA Polymerase	
3 step cycling				
Denaturation	15 s	95°C	Data collection at 60°C	
Annealing	15 s	60°C	Data collection at 60 C	
Extension	10 s	72°C		
Number of cycles	40			
Detection	Reporter	Excitation/emission	Channel	
Target	FAM	495/520 nm	Green	
Internal control	MAX	524/557 nm	Yellow	

ResultsThe isolation and identification of Salmonella were conducted according International Organization for Standardization (ISO) 6579-1 (2017). Depending to the traditional culture methods of stool samples the percentage of Salmonella spp. isolation was 72 (59.504%), (Table A-3). There was a significant difference (P<0.01) between the positive and negative results. The colonies of Salmonella spp. on XLD agar were small, circular, smooth, convex and red in color with black center (Fig.



Fig (B-1): Salmonella Colonies on XLD agar

Subsequently colonies on XLD agar was confirmed with biochemical tests revealed the inability of Salmonella to urea hydrolysis (Fig B-2), ability of Salmonella to ferment the glucose and produce the hydrogen sulfide gas when tested

on Triple Sugar Iron TSI media (Fig B-3) and Lysine Decarboxylation medium (LDC) To assist in the identification of Salmonellae (positive) Bacteria that are form Lysine Decarboxylase (LDC) in (Fig B-4).

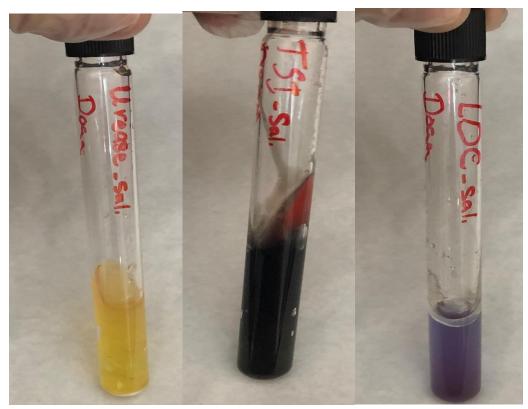


Fig (B-2): Urease test (-ve), Figure (B-3): TSI test (+ve) and Figure (B-4): LDC test (+ve)

To determine the presence or absence of salmonella pathogen based on amplification of *invA* gene using real time PCR, the sample considered to be positive if the corresponding fluorescence(FAM) curve lies clearly above the threshold line. The comparison between detection results of *Salmonella spp*. by traditional cultural methods and RT-PCR technique is described in

Table (A-3). Out of 72 cases (59.504%) that gave positive results as *Salmonella spp*. on culture media, out of 49 cases (40.495%) that gave negative results in culture and biochemical tests. While 68 cases (56.198%) were positive for PCR techniques and 53 cases (43.801%) were negative for PCR test.

Table (A-3): Percentages of Salmonella spp. isolation

Isolation Results	by culture		by PCR		X ² value
	Number	Percentage	Number	Percentage	$X^2 = 0.271$
Positive	72	59.504%	68	56.198%	Prob.= 0.602
Negative	49	40.495%	53	43.801%	
Total	121	100%	121	100%	

DISCUSSION

According to phenotypic characteristic, which is the main method for detection of bacteria. However, in some cases, PCR technique required for specific detection of Salmonella spp., particularly when atypical culture characteristics appear (Cohen et al., 1996, Daum et al., 2002). As well as these methods are not reliable for species designation due to morphological similarity among these organisms and the need of an expert person. Low sensitivity and specificity of these methods leads to controversy in interpretation of result. In vitro amplification of DNA by PCR method is a powerful tool in microbiological diagnostics (Malorny et al., 2003). Therefore, RT-PCR analysis with cultural methods were applied in this study to increase the detection specificity of Salmonella in Duhok city of Kurdistan Region and

Salmonellosis has remained one of the most common causes diarrhoeal diseases in human, and gastroenteritis is the typical disorder caused by non-typhoidal Salmonella infection as recorded by (Oliveiria et al., 2003). This high occurrence may be due to increase in the number of supermarkets and restaurants in Duhok province. These supermarkets are selling meat in parallel with other different products such as poultry, fish and meat. The result of present study is coincided with a study done by OWAIED, D.Y.H., (2016). They found that type S. typhimurium is more sovereignty in the studied species.

In most parts of the world, surveys have reported that S. Enteritidis and S. Typhimurium are the major serovars found in human (Saba et., al., 2013 and Antoine et., al. 2008). In Ghana and most other African countries, these types were also the most frequently isolated species from bloodstream infections (Evans et al., 2004 and Wilkens et al., 1997) and from diarrhoeal diseases (Bonkoungou et al., 2013), which is confirmed also by the findings of the present study.

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يوخته

پاشینه و ئارمانج:

سالمونیلا گەدەیی بەرپرسە ژ تووشبوونیّ دپتریا ھەودانیّن بەکتریا دھیّته قەگوھاستن ب خوارنیّ ل جیھانیّ .

ريك:

كارىّ قەكولىنا نوكە ھاتەكرن بو قەدىتن و نياسىنەكا بەشەكى يا سالمونىلا گەدەيى و قەدەركرى ژ نموونەيىّن پىساتيا وان نەخۆشان ئەوىّن ئامادەبووىن ل نەخۆشخانا ئازادى و ھىقى كو تووشى زك چوونىّ بووينە. ئەو قەدىتنى بكارئىنانا تەكنىكا كارلىّكا بەلمەرىّنى .

ئەنجام:

دناڤبەرا 121 نەخۆشان كو تووشى زك چوونى بووينە 72 حالەت (59,504 %) ژوان ئەنجامىن ئەرىنى يا سالمونىلا گەدەيى دان ئەوۋى ب رىكا چاندن ب چىنىن چاندنى. لى ب رىكا تاقىكرنىن كارلىكا بەلمەرىنا رىزبەند 68 حالەت (56,198 %) ژوان ئەنجامىن ئەرىنى يا سالمونىلا گەدەى دان. قى قەكولىنى 49 حالەت (40,495 %) ۋىلى شالمونىلا شوينگر ئەنجامىن نەرىنى دان ئەوۋى ب رىكا چاندنى و (53) حالەت (43,8 %) ۋىلى سالمونىلا شوينگر. ئەنجامىن نەرىنى دان ب رىكا تاقىكرنان ب تەكنىكا كارلىكا بەلمەرىنا رىزبەند. PCR دەرئەنجام:

قەكولىنا مە دياركر كو تەكنىكا RT – PCR يتر يا هوير و ھەستيارە ب بەراوردى دگەل رێكێن ئاسايى.

الخلاصة

الخلفية والأهداف: السالمونيلا المعوية هي المسؤولة عن التسبب في أكبر عدد من الالتهابات البكتيرية المنقولة بالغذاء في العالم. الطريقة: تم إجراء العمل البحثي الحالي للكشف والتعرف الجزيئي للسالمونيلا المعوية المعزولة من عينات البراز من المرضى الذين حضروا مستشفيي آزادي وهيفي يعانون من الإسهال باستخدام طريقة التقليديه والكشف باستخدام تقنيات تفاعل البلمرة .

النتائج: من بين 121 مريضاً أصيبوا بالإسهال ، 72 حالة (59.504٪) أعطت نتائج إيجابية للسالمونيلا المعوية بطريقة الزرع بالاطباق الزرعية، بينما من خلال اختبارات تفاعل البلمرة المتسلسل أعطت 68 حالة (56.198 ٪) بنائج إيجابية للسالمونيلا المعوية. هذه الدراسة ، (49) حالة (40.495 ٪) من غير السالمونيلا النيابة أعطت نتائج سلبية عن طريق الزرع و (53) حالة (43.8 ٪) من غير السالمونيلا النيابة. أعطى نتائج سلبية عن طريق اختبارات بتقنية تفاعل البلمرة المتسلسل

PCR

الاستنتاجات: أظهر بحثنا أن تقنية

RT-PCR

أكثر دقة وحساسية مقارنة بالطرق التقليدية