SPECIFIC GENES AFFECTING BODY WEIGHT IN IRAQI AWASSI SHEEP

YOUSIF M. S. AL-BARZINJI^{*} and ASO A. AMEEN^{**}

^{*}Dept. of Animal Resources, College of Agricultural Engineering Sciences, University of Salahaddin,

Kurdistan Region-Iraq

**Ministry of Agriculture & Water Resources-Kurdistan Region- Iraq

(Accepted for Publication: October 21, 2019)

ABSTRACT

The present study was to investigate genotypes of three genes (MSTN, CAST and MHC-DRB1) that related to body weight in Awassi lambs. A total of 52 lambs were used. The DNA concentration and DNA purity (A260/280) was calculated automatically by nanodrop and ranged between 10.53-53.94 ng/µl and 1.51-2.19, respectively. The RFLP-PCR results showed that the CAST and MHC-DRB1genes were polymorphism among individuals while the MSTN gene was monomorphism. Lambs with CC genotype for MHC-DRB1 locus have positive effect on body weight at most ages while lambs with AB genotype for CAST locus gave higher body weight at all ages and higher average daily gain. The lambs with BBCCBB genotypes for the three genes under study gave higher weight at weaning (22.15 kg/lamb) and higher average daily gain (0.197 kg/lamb/day) compared with all other genotypes. These results showed that Awassi lambs have potential for growth traits and the selection process with the aid of molecular technique can play a positive and rapid role for improvement this breed of sheep.

phenotypes.

KEYWORDS: RFLP-PCR, **MSTN**, **CAST**, **MHC-DRB1**, Weaning weight. <u>https://doi.org/10.26682/cajuod.2020.22.2.21</u>

INTRODUCTION

The native sheep breeds in Iraq are fattailed and carpet-wool type and are raised mainly for lamb and mutton production (Al-Rawi et al., 1996; Al-Azzawi et al., 1995; Al-Barzinji and Gardi, 2012 and 2015). The Awassi sheep, the most numerous small breed of Iraq (58.2%), ruminant are predominantly white with brown or red faces (FAO, 2000). Selection of farm animals depends on the availability of information's in animals. Classic breeding methodology by estimating breeding values requires a lot of efforts, time and fund. Identifying major genes can speed up (at more precise and accurate) genetic improvement of economic traits of sheep breeds. Identification and characterization of breeds is required to identify the genetic resources and also to prioritize breeds for conservation and development. Recently, the use of molecular markers, revealing polymorphism at the DNA level, has been playing an increasing role in animal genetics studies (Baumung et al., 2004).

Improvement of livestock has focused on the selection of breeding individuals with superior

increasingly advanced statistical methods that maximize selection for genetic gain, this simple approach has been extremely successful in increasing the quantity of agricultural output and productivity. However, information now available on the organization and functioning of the genome could be used in breeding programmers to improve a range of traits. Strategies to identify markers for traits and the application of these markers are described by reference to examples of loci that control a range of different traits (Williams, 2005). Therefore, the objective of the present study

the

development

of

With

is to identify the genetic polymorphism and best genotypes for lamb's growth using PCR-RFLP for three specific genes MSTN, CAST and MHC-DRB1.

MATERIALS AND METHODS Experimental Animals and Locations

This study was undertaken in commercial flock of Awassi sheep at the Bashblakh village / Kirkuk governorate (30 km east of Kirkuk). Ewes are exposed to rams for mating during end of May to the end of October, so that ewes

Y_Barzengy@Yahoo.com

^{*}E-Mail: Yousif.Noori@Su.Edu.Krd,

lambing occurred by end of October to end of November. A total of 52 ewes lambed out of 58 ewes mated, giving 52 lambs, six of them died during experiment. At lambing ewes and lambs were identified. Sexes, month of lambing were recorded. Ewes and lambs were raised together until weaning at 90 days of age. Male and female lambs were weighed at birth, one, two and three months of age.

Blood Collection and DNA Extraction

Whole blood was collected from each lamb at the end of the experiment through jugular vein into 5 ml Vacutainer tubes containing the anticoagulant, ethylenediaminetetra-acetic acid (EDTA). Samples were stored at -20°C until DNA extraction. All research works were carried out in both laboratory of Animal Molecular Genetics/College of Agriculture and Molecular Biology/Scientific Research Center/ Salahaddin University. Genomic DNA was isolated from samples using extraction kit (GeNet Bio, koria). The quantity and quality of DNA was checked by Nanodrop spectrophotometer and gel electrophoresis.

PCR Primers

The PCR programs were occurred out according to RFLP strategy portrayed by Davis *et al.*, (2000), Hanrahan *et al.*, (2004), Chu *et al.*, (2007) and Moradband *et al.*, (2011). Primers name, sequencing of primers with their restriction fragments are recorded in tables (1 and 2).

| | 、 / | , , | | U | |
|----------------|------------------|--------------------|------|------------------|---------------|
| Gene symbol | Primer sequences | | RE | Restriction site | Reference |
| | Forward | Reverse | | | |
| | | | | 5'C CGG 3' | |
| CAST | 5'-TGGGGC | 5'- GGTGGAG | Mspl | 3'GGC C 5' | Shahroudi et |
| | CCAATGACGCCATG | C AGCACTTC | | | al., (2011) |
| | ATG- 3' | TGATCACC-3' | | | |
| | | | | | |
| | | | | 5'TTT AAA 3' | Shahram and |
| MSTN | 5'-TGGCGTTA | 5´-AACAGCA | Dral | 3'AAA TTT 5' | Majid (2014) |
| | CTCAAAA | GTCAGCAG | | | |
| | GCAAA -3' | AGTCG-3 | | | |
| | | | | | |
| | | | | 5'GT AC 3' | |
| MHCDRB1 | 5'-TCTCTGCA | 5'-CTCGCCG | Rsal | 3'CA TG 5' | Ammer et al., |
| | GCACATTT | CTGCACAG TGAAAC-3' | | | (1992) |
| | CCTGG-3 | | | | |
| | | | | | |

 Table (1): Primers, Restriction Enzymes and restriction site for Candidate genes.

PCR Amplification

Thermo cycler (Applied **Biosystems**® Veriti® 96-Well Thermal Cycler Singapore) was conveyed in a final reaction volume of 20µl. A master mix for (46) samples for every gene was readied and an aliquot of 15 µl filled in every PCR tube. Five µl of DNA sample was added to each tube to make the last volume 20 µl [(A GoTaq blue Master Mix 10 µl, 25 units/ml Taq polemerase, each dNTPs is 200mM and MgCl2 is 1.5 mM), 2 µL RFLP primer (forward and reveres), 5µl (50ng) of DNA template and 3µl DNase free water)], (so as to accomplish homogeneity of reagents and decrease the risk of contamination, control reaction were situated up

without genomic DNA). DNA was amplified initial denaturation at 95°C for 5 min, followed by 30 cycles consisting of denaturation at 95°C for 30 sec, annealing 60°C for 1 min, extension at 72°C for 1 min, with final extension 72°C for 7 min. The amplification products were sizefractionated in a 2% agarose gel containing Ethidium Bromide in Tris-borate EDTA buffer and visualized under UV transillumination.

PCR – RFLP Analaysis

Following PCR amplification the digested PCR products with the appropriate restriction enzyme for each PCR products separately was carried out to identify if a mutation is present in a DNA samples or not. All digestion component requirements are shown in Table (2) Mixed delicately by pipetting, close the tube centrifuged for a few seconds in а microcentrifuge (Micro-12, Korea-Hanil Science Industrial). At that point the reactions were incubated for 2-4 hours at the pointed out temperature for the enzyme (generally 37 °C). after high temperature inactivation the product of the reaction digest, 10 µl of the digested samples were taken, included loading buffer, and were separated by electrophoreses on 2.5% agarose gel. One well (poket) every row was utilized for the length standard (100 bp DNA ladder) Bench top, volume 5 µl of length standard was utilized to gauge the size and concentration of the PCR product, the gel was visualized with Ethidum Bromide under the UV Transilluminator (Biostep-UST-20M-8K).

Statistic Analysis

The PROC GLM (General Linear Model) procedure SAS (2002) was used to analyze the data for the genotypes effects for weights at birth, one, two and three months (Weaning Weight) old and average daily gain from birth to weaning analyses were fitted to following equations:

$$Y_{ijklo} = \mu + A_i + S_j + C_k + P_l + \mathcal{E}_{ijklo}$$

Where: Y ijklo = Body weight at different age and daily of 0th lamb, of ith MSTN (Ai, i=1, BB), of jth CAST (Sj, j= 1, AB and j=2, BB), of kth MHC-DBR1 (Ck, k =1, AA, k=2, AB, k=3, AC and k=4, CC), of 1th all genes combinations (Pl, l=1,2,3,4,5 and 6), , μ = Population mean, \mathcal{E}_{ijklo} = random error. It was assumed to be normally and independently distributed with mean zero and variance $\delta^2 e$.

RESULTS AND DISCUSSION

Body weight of lambs at different ages

The overall mean of male and female lamb's body weights at birth, one, two, weaning weight and average daily gain from birth to weaning were gives in Table (3).

Table (2): PCR product digestion components with their volume for all genes under study.

| Digestion Component | Volume |
|-----------------------------------|--------------|
| Sterile, deionized water | 5.3 µl |
| Reaction 10X buffer | 2 µl |
| Acetylated BSA 10µg/µl | 0.2 µl |
| DNA(PCR product) mix by pipetting | 7 µl |
| Appropriate Reaction Enzyme | 0.5 µl (5 U) |
| Final Volume | 15 µl |

Molecular Profile

Limited molecular genetic information using RFLP markers is available on Iraqi Awassi sheep. Most of the results obtained in this study were the first attempt as a molecular characterization of this breed in Iraq. Fig. (1) shows extracted genomic DNA of total (46) blood samples from Awassi lamb which indicate that the quality of DNA samples are very good and can be used for future work and genetic analysis.

PCR- RFLP TECHNIQUE PCR-RFLP FOR CAST GENES

The PCR products for the CAST genes were digested with MspI restriction enzyme (2011) followed by all Nanekarani et al., samples run on 2.5% agarose gel electrophoresis to detect the genotype for each animal. The digested (622 bp) fragment in calpastatin gene gave two genotypes. The BB genotype exhibited 2 fragments of 336 and 286 bp and AB genotype exhibited 622,336 and 228 bp (Fig. 2). These results indicate that suitable diversity in Awassi sheep breed for calpastatin gene, that can used in future breeding programs. In the present study the diversity is perceived and consisting with reports on the other breeds (Gabor *et al.*, 2009 and Shahram and Majid, 2014). Allelic and

genotype frequency of CAST gene as given in Table (4).

| Table (3): Mean \pm SD of lamb's weight at different ages. | | | | |
|---|-----|-------------------|---------------------|-------------------------|
| Lamb | No. | Birth weight (Kg) | Weaning weight (Kg) | Average daily gain (Kg) |
| Male | 25 | 4.55 ± 0.68 | 21.38 ± 3.80 | 0.187 ± 0.03 |
| Female | 21 | 4.25 ± 0.69 | 19.82 ± 2.94 | 0.172 ± 0.03 |
| Overall | 46 | 4.4 ± 0.685 | 20.6 ± 3.37 | 0.179 ± 0.03 |



Fig. (1): DNA extraction image for 46 samples Awassi lambs C: Control without blood sample, M: DNA Ladder 100 *bp*



Fig. (2): PCR-RFLP analysis of 622 *bp* fragment of CAST gene by *MspI* enzyme on 2.5% agarose gel electrophoresis to separate bands with stained with 10µl Ethidium Bromide, C=Negative control and M= Molecular size.

| Genotypes | Allelic frequency | | Genotype frequency | |
|-----------|-------------------|------|--------------------|--------|
| BB(n=35) | В | А | BB | AB |
| AB(n=11) | 0.88 | 0.12 | 0.7609 | 0.2391 |

| Table (4): Allelic and Genotype frequency of CAST gene in | Awassi lambs |
|---|--------------|
|---|--------------|

PCR-RFLP for MSTN Gene

The *Dral* Restriction enzyme was used to digest the amplified PCR fragments (497 bp) according to Xianglong *et al.*, (2008). After that the digested product for each sample was run on 2.5% agarose gel electrophoresis to detect the genotype for all samples. The results show only one genotype for all samples, the BB genotype (427and 70 bp) was present in Fig. (3).

PCR-RFLP for MHC-DRB1 Gene

The PCR products for the MHC-DRB1 gene were digested with *RsaI* restriction enzyme (Ashrafi *et al.*, 2014). Samples were migrated on a 2.5 % agarose gel. Four genotypes were observed. Genotype (AB) with three fragments (279, 195 and 84 *bp*), genotype (CC) with two fragments (200 and 79 *bp*). The genotype (AC) with three fragments (279, 200 and 79*bp*) and (AA) with one fragment (279 *bp*), (Fig. 4). Allelic and Genotype frequency of MHC-DRB1 gene are demonstrated in Table (5).

Table (5): Allelic and Genotype frequency of MHC-DRB1 gene in Awassi lambs



Fig. (3): The results of analysis PCR-RFLP for MSTN gene by restriction enzyme *DraI* on 2.5 % agarose gel electrophoresis. Stained with 10µl Ethidium Bromide.C=Negative control and M= Molecular size.



Fig. (4): PCR-RFLP analysis of 279 *bp* fragment of MHC-DRB1 gene by *Rsal* enzyme on 2.5% agarose gel electrophoresis to separate bands with stained with 10µl Ethidium Bromide, C=Negative control and M= Molecular size marker (100 *bp* DNA ladder).

Molecular Markers and Quantitative Traits

The effect of genetic markers on growth traits in Awassi lambs under investigation study are showed in Fig. (5). The CC genotypes of MHC-DRB1 had a positive effect on body weight at all ages except body weight at birth and on average daily gain. The AB genotype at CAST locus gave higher body weight at all ages and higher daily gain compared with BB genotype. Thus the best genotype for lambs at three loci in the present study was BBCCBB which gave 22.15 kg/lambs with 0.197 kg/lamb/day (Table 5). These results reveal that selection can play major role to increase body weight especially weaning weight in Awassi sheep.

This result is similar with Khan *et al.*, (2012), who found in the CAST gene with MN genotype in Balkhi sheep give daily gain higher than the MM genotype at four months' age. Similar results were repoted by Gorlov *et al.*, (2016) for the CAST gene in Salsk sheep and by Ashrafi *et al.*, (2014) in Iranian Makuie sheep.

CONCLUSIONS

CAST and MHC-DRB1 genes were polymorphism while the MSTN gene was monomorphism. Lambs with CC genotype for MHC-DRB1 and lambs with AB genotype for CAST locus gave higher body weight at all ages and higher average daily gain. Lambs with BBCCBB genotypes gave about 1.5 kg/ lamb higher weight at weaning compared with all other genotypes. Thus, the investigation of three specific genes in Awassi lambs indicating the possibility of early selection of animals based on identifying specific genotype to improve the studied trait(s).

ACKNOWLEDGEMENTS

The authors are grateful to the Salahaddin University-Erbil to support this MS.C project study.



Fig. (5): Effect of loci genotype on the lambs body weight at different ages.

REFERENCES

- Al-Azzawi, W. A. R., Al-Salman, M. H., and Hamad, S.A. (1995).Relationship between body weight and dimention traits in Awassi sheep. Proceeding the Regional Symposiom on integrated Croplivestock system in west Asia and North Africa 6-8 November, Amman, Jordan; 284-290.
- AL-Barzinji, Y.M.S., and Gardi, H.E.A. (2012). Prediction of breeding value of awassi rams based on progeny performance in Kurdistan region of Iraq. Proceedings of the 15th AAAP Anim. Sci. Congress; 26-30.
- AL-Barzinji, Y.M.S., and Gardi, H.E.A. (2015). Genetic evaluation and factors affecting post weaning growth performance and body dimensions of Awassi lambs. J. of Zankoi Sulaimani;17 (3) (Part-A). P. 25-31.
- Al-Rawi, A. A, Al-Haboby A.H., and Salman A. (1996).Small ruminants breeding, reproduction physiology research as technology transfer in Iraq (W. Mourani, N. Haddad, Eds.), ICARDA West Asia Regional Program, Amman, Jordan.
- Ammer, H., Schwaiger, F.W., Kammerbauer, C., Gomolka, M., Arriens, A., Lazary, S., and Epplenj, T.(1992). Exonic polymorphism vs. intronic simple repeat hypervariability in

MHC-DRB genes. Immunogenetics; 35:.332-340.

- Ashrafi, F., Mardani, K., Ali, H.A., Darvishzadeh, R., and Farhadian, M. (2014). Association of molecular polymorphism in exon 2 of Ovar-DRB1 gene with weight traits in Iranian Makuie sheep breed. Turk J Vet Anim., Sci. ; 38:126-132.
- Baumung, R., Simianer, H.H., and Hoffmann, I. (2004). Genetic diversity studies in farm animals - a survey. J Anim Breed Genet; 121. P. 361–373.
- Chu, M. X., Liu, Z.H., Jiao, C.L., He, Y.Q., Fang, L., Ye, S.C., Chen, G.H., and Wang, J,Y.(2007). Mutation in BMPRIB and BMP15 genes are association with litter size in Small Tailed Han sheep (Ovis aries). J Anim Sci; 85:.598-603.
- Davis. G.H., Galloway, S.M., Ross, I.K., Gregen, S.M., Ward, J., Nimbkar, B.V., Ghalsasi, P.M., Nimbkar, C., *et al.* (2000). DNA tests in prolific sheep from eight countries provide new evidence on origin of the Booroola (FecB) mutation. Biol Reprod ; 66:1869 1874.
- FAO, (Food and Agriculture Organization), (2000). Quarterly bulletin of statistics 1. FAO. Rome, Italy.

*E-Mail: Yousif.Noori@Su.Edu.Krd,

Y_Barzengy@Yahoo.com

- Gabor, M.A., Trakovicha, A., and Miluchova, M. (2009). Analysis of polymorphism of CAST gene and Clpg gene in sheep by PCR-RFLP method. J. Zoot. Biotehnol; 42: 470-476.
- Gorlov, I., Shirokova, V., Randelin, A., Voronkova, V., Mosolova, N., Zlobina, E., etal. (2016). CAST/MspI gene polymorphism and its impact on growth traits of Soviet Merino and Salsk sheep breeds in the South European part of Russia. Turk J Vet Anim Sci.; 40:399-405.
- Hanrahan, J. P., Gregen, S. M., Mulsant, P., Mullen, M., Davis G. H., Powell R. and Galloways, S.(2004). Mutations in the genes for oocyte derived growth factors GDF9 and BMP15 are associated sheep (Ovis aries). Biol. Reprod. 70. P.900-909.
- Khan, S., Riaz, M., Ghaffar, A., and Khan, M. (2012). Calpastatin (CASTA) gene polymorphism and its association with average daily weight gain in Balkhi and Kajli sheep and Beetal Goat breeds. Pakistan J Zool ; 44(2): 377-382.
- Moradband, F., Rahimi, G., and Gholizadeh, M.(2011). Association of polymorphisms in Fecundity Genes of GDF9, BMP15 and BMP15-1B with Litter Size in Iranian Baluchi Sheep. Asian-Aust J Anim Sci ; 24 (9): 1179-1183.

- Nanekarani, S., Asadi, N., Khederzadeh, S.(2001). Genotypic frequency of calpastatin gene in lori sheep by PCR-RFLP method . 2nd International Conference on Environmental Science and Development IPCBEE vol.4, IACSIT Press, Singapore.
- SAS, Statistical Analyses System.(2002). SAS/STAT User's Guide, Version 8, first edition, Cary, NC, USA.
- Shahram, N. and Majid, G. (2014). Polymorphism of candidate genes for meat production in Lori sheep. IERI Procedia; 8: 18 – 23.
- Shahroudi, F. E., Nassiry, M. R., Valizadh, R., Moussavi, A.H., Tahmoorespour, M., and Ghiasi, H. (2006). Genetic polymorphism at MTNR1A, CAST and CAPN loci in Iranian Karakul sheep. Iranian J Biotechnology; 4(2): 117-122.
- Williams, J.L. (2005). The use of marker-assisted selection in animal breeding and biotechnology. Scientific and Technical Review, Office International Des Epizooties ; 24:379-391.
- Xianglong, L., Zhengzhu, I., Rongyan, L., Guiru, Z., Yuanfang, Z. G., and Lanhui, L. I. (2008). Deletion of TTTTA in 5 UTR of goat MSTN gene and its distribution and genetic effect on body weight. Front. Agric. China; 2(1): 103-109.

كارلێكردنى بۆھێڵە تايبەتەكان لەسەر كێۺ مەرى عەواسى عێراقى

پوخته

ئەم توێژىنەوەيە ئەنجام دراوە بۆ زانىنى كارلێكردنى سٽ بۆھێڵى تايبەت ,MSTN, CAST) (MHC-DRB1بە كێشى مەر لە مەرى عەواسىدا. پەنجاو دوو بەرخ بەكارھێنرا لە توێژىنەوەكەدا. چرى و پاكى دى ئێن ئەى ھەژماركرا بە بەكارھێنانى ئامێرى نانۆدرۆپ كە لە نێوان 10.53 بۆ ئەوەى نانۆگرام بۆ ھەموو مايكرۆليترێك وە 1.51 بۆ 2.19 بوو بەدواى يەكدا. ئەنجامى پى سى ئاړ ئەوەى دەرخست كەوا بۆھێڵى (CAST) وە (MHC-DRB1) جياوازبوون لە نێوان بەرخەكاندا وە بۆھێڵى بەرخانەى كەوا بۆھێڵى (MHC-DRB1) وە (MHC-DRB1) جياوازبوون لە نێوان بەرخەكاندا وە بۆھێڵى بەرخانەى جينۆتايپيان (AB) بوو لە بۆھێڵى (CC) بوولە بۆھێڵى (MHC-DRB1) وە ئەو بەرخانەى جينۆتايپيان (AB) بوو لە بۆھێڵى (CAST) كۆشيان زياتر بوو لەوانى تر. وە ئەو بەرخانەى كەوا جينۆتايپيان (BBCCB8) يە بۆ ھەر سٽ بۆھێڵەكان بەرزترين كێشيان داوە لە كاتى لەشيرگرتنەوە كە 2.15 كىلۆگرام بوو بۆ ھەر بەرخێك وە بەرزترين زيادكردنى رۆژانەيان ھەبوو كە 2.19 كىلۆگرام بوو بۆ ھەر بەرخێك لە يۆۋێكدا.ئەم ئەنجامانە ئەوە دەرئەخەن كەوا مەرى كە 1.19 كىلۆگرام بوو بۆ ھەر بەرخيلە مەيزەتلەم ئەنجامانە ئەۋە دەرئەخەن كەۋا مەرى كە 1.19 كىلۆگرام بوو بۆھەر بەرخينى ھەنجامانە ئەيە دەرئەخەن كەۋا مەرى كە 1.19 كىلۆگرام بوو بۆھەر بەرخيك لە يرۆزىكوا.ئەم ئەنجامانە ئەيە دەرئەخەن كەوا مەرى مەرەلىي تواناييەكى باشى گەشەكردنى ھەيەو ئەنجام دانى پرۆسەي ھەلبژاردن بە بەكارھێنانى مەرەرە لە عزرامەرونى بۇرلۇلىرى يۆزەتىغانەو خىراى ئەبىت بۆ بەرەوپىشچوونى ئەم جۆرە

تاثير الجينات الخاصة على وزن الجسم في الأغنام العواسية العراقية

الخلاصة

أجريت هذه الدراسة لمعرفة تأثير ثلاث جينات خاصة بالأوزان (MSTN, CAST, MHC-DRB1) في الحملان العواسية. أستخدمت 52 حملا في الدراسة. قيست تركيز و نقاوة الدنا باستخدام جهاز نانودروب وتراوح القيم مابين 10.53 الى 53.94 نانوغرام لكل ملليتر و 1.51 لى 2.19 على التوالي. أوضحت نتائج جهاز البلمرة الحرارية بأن كل من (CAST) و (MHC-DRB1) أعطت حزم مختلفة مابين الحملان ولكن الموقع الجيني (MSTN) أعطت حزمة واحدة لكل الحيوانات تحت الدراسة. الحملان دو تركيب وراثي (CC) للجين (MSTN) أعطت حزمة واحدة لكل الحيوانات تحت الدراسة. أعطوا أعلى الأوزان والزيادة الوزنية اليومية عند الفطام. وعلى نطاق المواقع الثلاث فأن ألتركيب الوراثي (BBCCBB) قد أعطت أعلى معدل لوزن الفطام 22.15 كغم لكل حمل وأعلى معدل للزيادة الوزنية اليومية من الولادة للفطام والبالغ 20.19 كغم لكل حمل وأعلى معدل للزيادة الوزنية اليومية من الولادة للفطام والبالغ 10.57 كغم لكل حمل وأعلى معدل للزيادة الوزنية اليومية من الولادة للفطام والبالغ 10.57 كغم لكل حمل وأعلى معدل للزيادة الوزنية اليومية من الولادة للفطام والبالغ 10.57 كغم لكل حمل وأعلى معدل للزيادة الوزنية اليومية من الولادة للفطام والبالغ 10.57 كغم لكل حمل وأعلى التركيب الوراثي والزيادة الولادة للفطام والبالغ 10.57 كغم لكل حمل يوميا. هذه النتائج معدل للزيادة الوزنية اليومية من الولادة للفطام والبالغ 10.57 كغم لكل حمل يوميا. هذه النتائج معدل للزيادة الوزنية اليومية من الولادة للفطام والبالغ 10.57 كغم لكل حمل يوميا. هذه النتائج معدل للزيادة الوزنية اليومية من الولادة للفطام والبالغ 10.57 كغم لكل حمل يوميا. هذه النتائج تشير الى أن الحملان العواسية لها قدرة وراثية جيدة للنمو وأن الأنتخاب على أساس التقنيات الوراثية الجزيئية لها دور فعال في تسريع عمليات التحسين للأوزان في الأغنام العواسية العراقية.